

Illinois Institute of Technology

Radiation Biophysics Lecture 13: Genetics Andrew Howard

08/07/2008

RadBio Bootcamp: Lecture 13

p. 1 of 26

Plan for this lecture

- ◆ Genetics
 - Chromosome Breakage
 - . Single hits
 - . Double hits
 - Gene mutations
 - . Frameshifts
 - . Introns
 - Molecular Biology 102

08/07/2008

RadBio Bootcamp: Lecture 13

p. 2 of 26

Generalizations about Genetics

- ◆ Main focus here is on ways that ionizing radiation can damage the physical and replicative properties of DNA
- ◆ We'll look at three length scales:
 - The individual base-pair (~0.5 nm)
 - The gene (1000 bp - many nm)
 - The entire chromosome (200 nm or more)
- ◆ Focusing only on single-base mutations (substitutions, deletions, insertions) does not tell the whole story

08/07/2008

RadBio Bootcamp: Lecture 13

p. 3 of 26

Early studies

- ◆ T.H. Morgan studied chromosomes in *Drosophila*
- ◆ *Tradescantia* (spiderwort) microspores: Sax, 1938-1950
- ◆ More *Tradescantia*: Lea et al, 1946: Provided quantitative data on chromosomal aberrations during first mitotic cycle



08/07/2008

RadBio Bootcamp: Lecture 13

p. 4 of 26

Chromosome Breakage

- ◆ Quite common result of radiation exposure
- ◆ Can happen before replication: then the structural defect will be replicated (if the cell survives)
- ◆ Can happen afterward: then one of the two chromatids will differ from the other one.
- ◆ Types of damage: Subchromatid, Chromatid, Chromosome
- ◆ Repair tends to be rapid

08/07/2008

RadBio Bootcamp: Lecture 13

p. 5 of 26

Single-Hit Breaks

- ◆ Cartoons show * as centromere and ^ as break
- ◆ Single-Hit:
 $A B C * D E F ^ G H I \rightarrow A B C * D E F + G H I$
- ◆ This can recombine in a number of ways:
 $A B C * D E F I H G$
 $I H G A B C * D E F$
 $G H I A B C * D E F$
- ◆ ... or a circle plus a fragment without a centromere, which won't be able to use the spindle machinery to get sorted.

08/07/2008

RadBio Bootcamp: Lecture 13

p. 6 of 26

Double Hits

- ◆ $A B C * D \wedge E F \wedge G H I \rightarrow$
 $A B C * D + E F + G H I$
- ◆ Numerous ugly combinations can arise, e.g.
 $E F A B C * D I H G$
- ◆ Acentrics can readily arise

08/07/2008

RadBio Bootcamp: Lecture 13

p. 7 of 26

Multiple Hits in Replicated Chromosomes

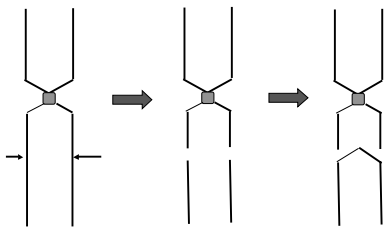
- ◆ See figs. 13.1 and 13.2
- ◆ Major losses of genetic information possible
- ◆ Balanced Translocations: no harm done!
- ◆ Dicentrics--two centromeres in one pair of chromosomes.
- ◆ Sister chromatid exchange: exchange of DNA fragments from one chromatid to another between the two chromatids of a single chromosome
 - Important for chemical mutagens
 - Not common with radiation

08/07/2008

RadBio Bootcamp: Lecture 13

p. 8 of 26

Double-stranded breaks

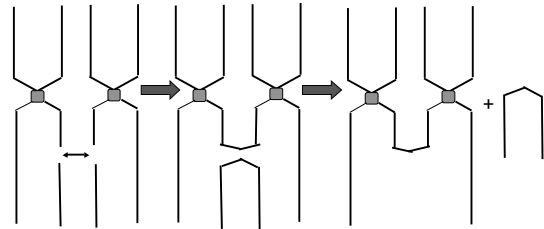


08/07/2008

RadBio Bootcamp: Lecture 13

p. 9 of 26

Recombination



08/07/2008

RadBio Bootcamp: Lecture 13

p. 10 of 26

Breakage and Exchange Hypotheses

- ◆ Breakage hypothesis:
Types of aberrations that require one break follow linear dose-response; those requiring two or three follow quadratic or cubic dose-response.
- ◆ Exchange hypothesis:
emphasizes reciprocal exchanges among chromosomal materials
Data don't support this one much.

08/07/2008

RadBio Bootcamp: Lecture 13

p. 11 of 26

Gene Mutations

- ◆ We've discussed these in detail previously
- ◆ Types:
 - Deletions (frameshift)
 - Additions (slightly less likely) (frameshift)
 - Substitutions (e.g. C for T; no frameshift)
- ◆ Remember that three bases code for an amino acid!
- ◆ If we skip a single base, we can throw off every single amino acid that is coded for downstream of the error.
- ◆ Mutation frequencies are often linear with dose

08/07/2008

RadBio Bootcamp: Lecture 13

p. 12 of 26

Effect of Frameshift

- ◆ Suppose the DNA duplex looks like this:
 5'- A - T - C - G - C - A - A - 3'
 3'- T - A - G - C - G - T - T - 5'
- ◆ Now we delete one base, so that the coding strand is
 3'- T - A - G - G - T - T - 5'
- ◆ Now the amino acids coded for will be different.

08/07/2008 RadBio Bootcamp: Lecture 13 p. 13 of 26

Frameshifts (continued)

- Most common form of (local) DNA damage
- Defined as any form of DNA damage that leads to a loss of one base during replication or transcription.
- Chemistry
 - Deletion of bases
 - Fragmentation of sugar-phosphate backbone
 - Distortion of base-base hydrogen bonds and other 3-D elements
- Results in complete misreading of remainder of message

08/07/2008 RadBio Bootcamp: Lecture 13 p. 14 of 26

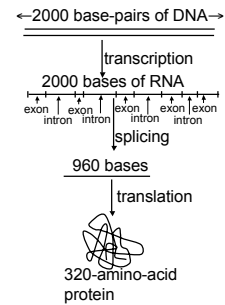
Genes

- ◆ A gene is a unit within a chromosome (i.e., a stretch of DNA) that codes for a specific function.
- ◆ Most genes ultimately code for proteins; *but*
- ◆ Some code for transfer RNAs
- ◆ Eukaryotic genes often have exons and introns:
 - Exons are segments of a gene that are ultimately turned into proteins
 - Introns are segments of a gene that are removed from the message before translation

08/07/2008 RadBio Bootcamp: Lecture 13 p. 15 of 26

How Introns are Removed

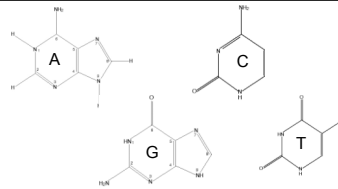
- ◆ Suppose we begin with a 2000-base-pair gene.
- ◆ It will be transcribed to make a 2000-base messenger RNA molecule
- ◆ That gets chopped up to make up a ~960 base ultimate message
- ◆ This will produce a 320-amino acid protein (960/3 = 320)



08/07/2008 RadBio Bootcamp: Lecture 13 p. 16 of 26

Structural components of genes

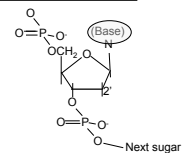
- ◆ Bases
 - Adenine
 - Cytosine
 - Guanine
 - Thymine
- ◆ Phosphate-deoxyribose backbone
- ◆ Double helix enables the replication and transcription processes and offers physical stabilization



08/07/2008 RadBio Bootcamp: Lecture 13 p. 17 of 26

Review of DNA backbone

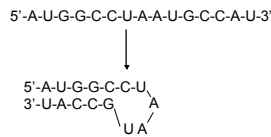
- ◆ Sugar-phosphate backbone on each strand has a nitrogenous acid base attached at the 1' position of the ribose ring
- ◆ The lack of an -OH group on the 2' position prevents formation of a reactive intermediate; this makes DNA more stable than RNA
- ◆ DNA's double-helical structure also helps its stability.



08/07/2008 RadBio Bootcamp: Lecture 13 p. 18 of 26

RNA hairpins

- ◆ DNA is almost always double helical; Watson-Crick base pairing contributes to stability
- ◆ RNA sometimes has complementary bases that allow for bases to pair up around a (hairpin) loop



08/07/2008

RadBio Bootcamp: Lecture 13

p. 19 of 26

Special features

- ◆ Code is redundant
- ◆ $4^3 = 64$ codons
- ◆ Only 20 amino acids plus a few control codes; most amino acids have more than one codon each associated with them.
- ◆ Middle base is generally conserved; all the codons for any given amino acid generally have the same middle base.
- ◆ First RNA base may be sloppy

08/07/2008

RadBio Bootcamp: Lecture 13

p. 20 of 26

A problem to ponder

- ◆ Explain why a frameshift of 3 bases is less likely to be fatal than a frameshift of 1 or 2 bases.

08/07/2008

RadBio Bootcamp: Lecture 13

p. 21 of 26

Errors in Fidelity and Their Consequences

- ◆ How do chemicals & radiation affect fidelity (rate of mutation)?
- ◆ Increase likelihood of replication error
- ◆ Radiation: bond disruption in bases (or sugars)
 - Direct (radiation breaks bonds in DNA)
 - Indirect (radiation causes free radicals; free radicals break bonds in DNA)

08/07/2008

RadBio Bootcamp: Lecture 13

p. 22 of 26

Chromosomal Instability

- ◆ The observation is that cell death continues for several generations after the initial exposure
- ◆ The explanation is that the chromosome becomes unstable, i.e. it loses some of its ability to remain structurally sound.
- ◆ This instability is heritable and therefore accounts for some of these long time-scale effects
- ◆ Little (1990): decreases in cloning efficiency lasts for 20-30 population doubling times in mammalian cells

08/07/2008

RadBio Bootcamp: Lecture 13

p. 23 of 26

Results in intact eukaryotes

- ◆ Not as easy to study as in culture
- ◆ Most work historically has been done in *Drosophila*
 - Rate of production of lethal mutations is linear with dose
 - Dose rate is irrelevant
 - Male gamete's sensitivity varies greatly with the stage of development at which irradiation occurs
 - Rate of mutations ~ $1.5 \cdot 10^{-6} - 8 \cdot 10^{-8}$ per locus per cGy.
- ◆ n.b.: a locus is a testable genetic element; it's typically one gene, although it could be a group of genes controlled by a single promoter

08/07/2008

RadBio Bootcamp: Lecture 13

p. 24 of 26

Results in Mammals

- ◆ Concern was raised in the 1960's that we shouldn't base mammalian exposure risks on *Drosophila* data
- ◆ Mouse specific locus test: suggests
 - 2.2×10^{-7} mutations/locus/cGy.
 - Dose-rate matters: fewer observed mutations at lower dose rate.
- ◆ Difficult to extrapolate these mouse results to humans
- ◆ Rates around $1 - 5 \times 10^{-7}$ is typical.

08/07/2008

RadBio Bootcamp: Lecture 13

p. 25 of 26

Repair

- ◆ Cell has mechanisms to recognize & replace faulty bases before they have a chance to be replicated
- ◆ Some injury may disrupt a large enough segment of DNA that repair either fails or is error-prone

08/07/2008

RadBio Bootcamp: Lecture 13

p. 26 of 26